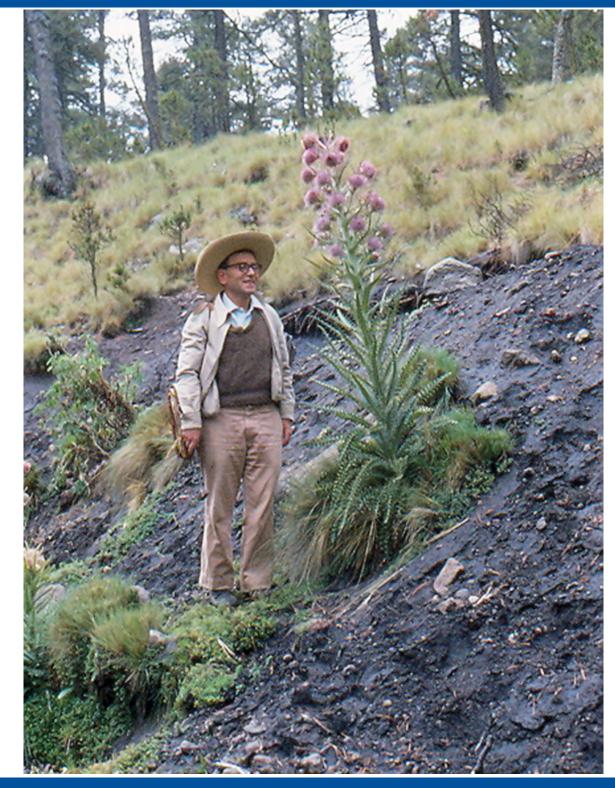


# Polib Tánica

# ISSN 1405-2768





Julio 2023







**Julio 2023** 





Instituto Politécnico Nacional "La Técnica al Servicio de la Patria"







# PÁG.

## CONTENIDO

- 1 Riqueza y distribución altitudinal de hepáticas epifitas del bosque mesófilo de montaña de Hidalgo, México Richness and altitudinal distribution of epiphytic liverworts from the cloud forest of Hidalgo, Mexico José Francisco Juárez López | Arturo Sánchez González | Maritza López Herrera | Dorismilda Martínez Cabrera
- 17 Descripción e ilustración del desarrollo morfogénico de los gametófitos y esporófitos jóvenes de Asplenium blepharophorum Bertol. (Aspleniaceae-Polypodiidae) en tres sustratos naturales Description and illustration of the morphogenic development of the young gametophytes and sporophytes of Asplenium blepharophorum Bertol. (Aspleniaceae-Polypodiidae) in three natural substrates Adriana Rojas Cano | María de la Luz Arreguín Sánchez | Rafael Fernández Nava | David Leonor Quiroz García
- 39 Análisis morfométrico de Agave sensu stricto (Asparagaceae: Agavoideae) en Veracruz y áreas adyacentes de Puebla, México Morphometric analysis of Agave sensu stricto (Asparagaceae: Agavoideae) in Veracruz and adjacents areas of Puebla, Mexico Carlos Rafael Arzaba Villalba | Miguel Cházaro Bazáñes | Mario Luna Cavazos | Edmundo García Moya
- 61 Variación clinal de caracteres fenotípicos y fisiológicos en *Pinus hartwegii* Lindl., para la Estación Forestal Experimental Zoquiapan, México *Clinal variation of phenotypic and physiological characters in* Pinus hartwegii Lindl., *for the Zoquiapan Experimental Forest Station, Mexico* Adrián López López | María Isabel Palacios Rangel | Cuauhtémoc Sáenz Romero | Villanueva Morales Antonio | Victoria Pacheco Almaraz
- 81 Composición, estructura y estado de la regeneración arbórea en un gradiente altitudinal en un bosque templado de Guadalupe y Calvo, Chihuahua Composition, structure and status of tree regeneration in an altitudinal gradient in a temperate forest of Guadalupe y Calvo, Chihuahua Samuel Alberto García García | Eduardo Alanís Rodríguez | Óscar Alberto Aguirre Calderón | Eduardo Javier Treviño Garza | Luis Gerardo Cuéllar Rodríguez | Alejandro Collantes Chávez Costa
- 101 Efectos del manejo forestal en la emisión de CO<sub>2</sub> de un suelo umbrisol en bosques de Durango, México Effects of forest management on the soil CO<sub>2</sub> emission of an umbrisol in forests from Durango, Mexico Erik Orlando Luna Robles | Israel Cantú Silva | Francisco Javier Hernández | Silvia Janeth Bejar Pulido
- 115 Influencia del conocimiento ecológico tradicional y la altitud en la estructura y diversidad arbórea de los cercos vivos del corredor biológico Chichinautzin, México Influence of traditional ecological knowledge and altitudinal gradient on richness, structure and tree diversity of live fences in the Chichinautzin biological corridor, Mexico Emir Basurto García | Hortensia Colin Bahena | Rafael Monroy Ortiz | Alejandro García Flores | Leonardo Beltrán Rodríguez
- Efecto del medio de cultivo y escotoperiodo en la germinación de semillas y crecimiento *in vitro* de *Guarianthe skinneri* (Bateman) Dressler & W.E. Higgins (Orchidaceae)
   Effect of culture media and skotoperiod on the germination of seeds and growth in vitro of Guarianthe skinneri (Bateman) Dressler & W.E. Higgins (Orchidaceae)
   Fabiola Hernández Ramírez | Leobardo Iracheta Donjuan | Anne Damon | Sylvia Patricia Fernández Pavía | Karina Guillén Navarro
- Plant regeneration from indirect somatic embryogenesis of Agave salmiana Otto ex Salm-Dyck subsp. salmiana using zygotic embryo obtained by incasa pollination as explants
   Regeneración de plantas por embriogénesis somática indirecta de Agave salmiana Otto ex Salm-Dyck subsp. salmiana usando embriones cigóticos obtenidos por polinización In-casa como explantes
   Blanca Vianey Angeles Vázquez | Jorge Alvarez Cervantes | Xóchitl Tovar Jiménez | Benjamín Rodríguez Garay
- 183 Flavonoids, anthocyanins and total macronutrients in handmade products of blackberry (*Rubus* sp.) from Atecaxil, Veracruz, Mexico *Flavonoides, antocianinas y macronutrientes totales en productos artesanales de zarzamora* (Rubus *sp.) de Atecáxil, Veracruz, México* Vianey del Rocio Torres Pelayo
- 203 Composición química, actividad antioxidante, antiinflamatoria y antiproliferativa del extracto de callos derivado de Acalypha californica Bentham Chemical composition, antioxidant, antiinflammatory an antiproliferative activity of callus extract derived from Acalypha californica Bentham Lesyanny Hechavarría Pérez | Luisa Alondra Rascón Valenzuela | Armando Tejeda Mansir | José Alberto Pérez Burgos | Gloria Irma Ayala Astorga
- 225 Dinámica de la Etnobotánica médica de los pobladores de Córdoba, Argentina. Aportes de la Encuesta Nacional de Folklore (1921) a la comprensión de los cambios en el uso y percepción de plantas medicinales Dynamics of the medical Ethnobotany of the inhabitants of Córdoba, Argentina. Contributions of the National Survey of Folklore (1921) to understanding changes in the use and perception of medicinal plants Cecilia Trillo | Bárbara Arias Toledo
- 249 Ethnobotany of medicinal plants Used for healers of the Chol ethnic group from Tila, Chiapas, Mexico Etnobotánica de plantas medicinales usadas por curanderos del grupo étnico Chol de Tila, Chiapas, México José del Carmen Rejón Orantes | Sabina Andrea Sánchez-Cartela | Wilbert Gutiérrez-Sarmiento | Oscar Farrera-Sarmiento | Miguel Pérez de la Mora
- 265 Estudio de plantas medicinales utilizadas en San José Iturbide, Guanajuato, México Study of medicinal plants used in San Jose Iturbide, Guanajuato, Mexico Eduardo Alberto Lara Reimers | Carlos Omar Hernández Robledo | Pablo Preciado Rangel | Oscar Sariñana Aldaco
- 287 Percepción y significados del guaje rojo Leucaena esculenta (DC.) Benth. en la cultura ngiwa de Puebla, México Perception and meanings of the red guaje Leucaena esculenta (DC.) Benth. in the ngiwa culture from Puebla, Mexico Guadalupe García Maceda | Arturo Hernández Montes | María Carmen Ybarra Moncada | Rocío Guadalupe Casañas Pimentel
- 311 Intergenerational transmission of traditional ecological knowledge about medicinal plants in a riverine community of the Brazilian Amazon Transmisión intergeneracional del conocimiento ecológico tradicional sobre las plantas medicinales en una comunidad ribereña de la Amazonía Brasileña Rogério Lima Mota | Iani Dias Lauer-Leite | Jaílson Santos de Novais

# POLIB TÁNICA

Núm. 56

ISSN electrónico: 2395-9525

Julio 2023



### Portada

Jerzy Rzedowski Rotter (1926-2023). Considerado uno de los botánicos más influyentes de México. Incursionó en diversas disciplinas botánicas como taxonomía, florística, fitogeografía y ecología. Formó varios herbarios institucionales y recolectó muestras de la flora mexicana, logrando una colección que superó los 50,000 ejemplares. Trabajó en la Flora Fanerogámica del Valle de México y en la Flora del Bajío y Regiones Adyacentes; también escribió el libro La Vegetación de México, obra clásica de la literatura botánica mexicana. A lo largo de su carrera, se dedicó además a la enseñanza y formación de botánicos. Su obra incluye la publicación de 7 libros, 47 capítulos de libros, 128 artículos en revistas científicas y 31 fascículos de floras. Descubrió alrededor de 190 nuevas especies de plantas mexicanas y más de 85 especies de hongos, plantas y animales mexicanos recibieron su nombre en su honor.

Jerzy Rzedowski Rotter (1926-2023). Considered one of the most influential botanists in Mexico. He ventured into several botanical disciplines such as taxonomy, floristics, phytogeography, and ecology. He formed several institutional herbaria, and collected samples of Mexican flora, achieving a collection that exceeded 50,000 numbers. He worked on the Phanerogamic Flora of the Valley of Mexico and the Flora of the Bajio and Adjacent Regions; he also wrote the book The Vegetation of Mexico, a classic work of Mexican botanical literature. Throughout his career, he was also dedicated to teaching and training botanists. His work includes the publication of 7 books, 47 book chapters, 128 articles in scientific journals, and 31 fascicles of floras. He discovered about 190 new species of Mexican plants and more than 85 species of Mexican fungi, plants, and animals were named in his honor.

# por/by Rafael Fernández Nava





# INSTITUTO POLITÉCNICO NACIONAL

Director General: *Dr. Arturo Reyes Sandoval* Secretario General: *Ing. Arq. Carlos Ruiz Cárdenas* Secretario Académico: *Mtro. Mauricio Igor Jasso Zaranda* Secretario de Innovación e Integración Social: *M. en C. Ricardo Monterrubio López* Secretario de Investigación y Posgrado: *Dra. Laura Arreola Mendoza* Secretario de Servicios Educativos: *Dra. Ana Lilia Coria Páez* Secretario de Administración: *M. en C. Javier Tapia Santoyo* Director de Educación Superior: *Dra. María Guadalupe Ramírez Sotelo* 

# ESCUELA NACIONAL DE CIENCIAS BIOLÓGICAS

Director:

Dr. Isaac Juan Luna Romero Subdirectora Académica: M. en C. Martha Patricia Cervantes Cervantes Jefe de la Sección de Estudios de Posgrado e Investigación: Dr. Gerardo Aparicio Ozores Subdirector de Servicios Educativos e Integración Social: Biól. Gonzalo Galindo Becerril

**POLIBOTÁNICA**, Año 28, No. 56, julio 2023, es una publicación semestral editada por el Instituto Politécnico Nacional, a través de la Escuela Nacional de Ciencias Biológicas. Unidad Profesional Lázaro Cárdenas, Prolongación de Carpio y Plan de Ayala s/n, Col. Santo Tomas C.P. 11340 Delegación Miguel Hidalgo México, D.F. Teléfono 57296000 ext. 62331. http://www.herbario.encb.ipn.mx/, Editor responsable: Rafael Fernández Nava. Reserva de Derechos al Uso Exclusivo del Título No. 04-2015-011309001300-203. ISSN impreso: 1405-2768, ISSN digital: 2395-9525, ambos otorgados por el Instituto Nacional del Derecho de Autor. Responsable de la última actualización de este número, Unidad de informática de la ENCB del IPN, Rafael Fernández Nava, Unidad Profesional Lázaro Cárdenas, Prolongación de Carpio y Plan de Ayala s/n, Col. Santo Tomas CP 11340 Delegación Miguel Hidalgo México, D.F.

Las opiniones expresadas por los autores no necesariamente reflejan la postura del editor de la publicación.

Queda estrictamente prohibida la reproducción total o parcial de los contenidos e imágenes de la publicación sin previa autorización del Instituto Politécnico Nacional.

#### REVISTA BOTÁNICA INTERNACIONAL DEL INSTITUTO POLITÉCNICO NACIONAL

#### **EDITOR EN JEFE**

Rafael Fernández Nava

#### **EDITORA ASOCIADA**

María de la Luz Arreguín Sánchez

#### **COMITÉ EDITORIAL INTERNACIONAL**

Christiane Anderson University of Michigan Ann Arbor. Michigan, US

*Heike Vibrans* Colegio de Postgraduados Estado de México, México

Graciela Calderón de Rzedowski Instituto de Ecología del Bajío Páztcuaro, Mich., México

> *Theodore S. Cochrane* University of Wisconsin Madison, Wisconsin, US

Hugo Cota Sánchez University of Saskatchewan Saskatoon, Saskatchewan, Canada

*Fernando Chiang Cabrera* Universidad Nacional Autónoma de México Ciudad de México, México

> Thomas F. Daniel California Academy of Sciences San Francisco, California, US

Francisco de Asis Dos Santos Universidad Estadual de Feira de Santana Feira de Santana, Brasil

> Carlos Fabián Vargas Mendoza Instituto Politécnico Nacional Ciudad de México, México

José Luis Godínez Ortega Universidad Nacional Autónoma de México Ciudad de México, México

> José Manuel Rico Ordaz Universidad de Oviedo Oviedo, España

*Edith V. Gómez Sosa* Instituto de Botánica Darwinion Buenos Aires, Argentina

Jorge Llorente Bousquets Universidad Nacional Autónoma de México Ciudad de México, México

> Delia Fernández González Universidad de León León, España

Jerzy Rzedowski Rotter Instituto de Ecología del Bajío Pátzcuaro, Mich., México

Luis Gerardo Zepeda Vallejo Instituto Politécnico Nacional Ciudad de México, México

*Claude Sastre* Muséum National d'Histoire Naturelle París, Francia

> Mauricio Velayos Rodríguez Real Jardín Botánico Madrid, España

Noemí Waksman de Torres Universidad Autónoma de Nuevo León Monterrey, NL, México

> Julieta Carranza Velázquez Universidad de Costa Rica San Pedro, Costa Rica

> > *Tom Wendt* University of Texas Austin, Texas, US

#### DISEÑO Y FORMACIÓN ELECTRÓNICA Luz Elena Tejeda Hernández

**OPEN JOURNAL SYSTEM Y TECNOLOGÍAS DE LA INFORMACIÓN** *Pedro Aráoz Palomino*  Toda correspondencia relacionada con la revista deberá ser dirigida a:

Dr. Rafael Fernández Nava Editor en Jefe de



Departamento de Botánica Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional Apdo. Postal 17-564, CP 11410, Ciudad de México

> Correo electrónico: polibotanica@gmail.com rfernan@ipn.mx

Dirección Web http://www.polibotanica.mx

POLIBOTÁNICA es una revista indexada en:

CONAHCYT, índice de Revistas Mexicanas de Investigación Científica y Tecnológica del Consejo Nacional de Humanidades, Ciencia y Tecnología.

SciELO - Scientific Electronic Library Online.

Google Académico - Google Scholar.

DOAJ, Directorio de Revistas de Acceso Público.

Dialnet portal de difusión de la producción científica hispana.

REDIB Red Iberoamericana de Innovación y Conocimiento Científico.

LATINDEX, Sistema regional de información en línea para revistas científicas de América Latina, el Caribe, España y Portugal.

PERIODICA, Indice de Revistas Latinoamericanas en Ciencias.





Polibotánica ISSN electrónico: 2395-9525 polibotanica@gmail.com Instituto Politécnico Nacional México http://www.polibotanica.mx

FLAVONOIDS, ANTHOCYANINS AND TOTAL MACRONUTRIENTS IN HANDMADE PRODUCTS OF BLACKBERRY (*Rubus* sp.) FROM ATECAXIL, VERACRUZ, MEXICO

# FLAVONOIDES, ANTOCIANINAS Y MACRONUTRIENTES TOTALES EN PRODUCTOS ARTESANALES DE ZARZAMORA (*Rubus* sp.) DE ATECÁXIL, VERACRUZ, MÉXICO

Felipe-Mendoza, Neil Abdiel; María de Jesús Martínez-Hernández; María del Carmen Ramírez-Benítez; Karla Daniela Hernández-González; Jorge Molina-Torres; Mayvi Alvarado-Olivarez y Vianey del Rocío Torres-Pelayo

FLAVONOIDS, ANTHOCYANINS AND TOTAL MACRONUTRIENTS IN HANDMADE PRODUCTS OF BLACKBERRY (*Rubus* sp) FROM ATECAXIL, VERACRUZ, MEXICO

FLAVONOIDES, ANTOCIANINAS Y MACRONUTRIENTES TOTALES EN PRODUCTOS ARTESANALES DE ZARZAMORA (*Rubus* sp) DE ATECÁXIL, VERACRUZ, MÉXICO

POLIB®TÁNICA

Instituto Politécnico Nacional

Núm. **56**: 183-201 México. Julio 2023 DOI: 10.18387/polibotanica.56.10



Este es un artículo de acceso abierto bajo la licencia Creative Commons 4.0 Atribución-No Comercial (<u>CC BY-NC 4.0 Internacional</u>).

# Flavonoids, anthocyanins and total macronutrients in handmade products of blackberry (*Rubus* sp) from Atecaxil, Veracruz, Mexico

#### Flavonoides, antocianinas y macronutrientes totales en productos artesanales de zarzamora (*Rubus* sp) de Atecáxil, Veracruz, México

Felipe-Mendoza, Neil Abdiel; María de Jesús Martínez-Hernández; María del Carmen Ramírez-Benítez; Karla Daniela Hernández-González; Jorge Molina-Torres; Mayvi Alvarado-Olivarez y Vianey del Rocío Torres-Pelayo

FLAVONOIDS, ANTHOCYANINS AND TOTAL MACRONUTRIENTS IN HANDMADE PRODUCTS OF BLACKBERRY (*Rubus* sp) FROM ATECAXIL, VERACRUZ, MEXICO

FLAVONOIDES, ANTOCIANINAS Y MACRONUTRIENTES TOTALES EN PRODUCTOS ARTESANALES DE ZARZAMORA (*Rubus* sp) DE ATECÁXIL, VERACRUZ, MÉXICO

### POLIB®TÁNICA

Instituto Politécnico Nacional

Núm. 56: 183-201. Julio 2023

DOI: 10.18387/polibotanica.56.10

#### Neil Abdiel Felipe-Mendoza

Facultad de Biología, Universidad Veracruzana, Circuito Gonzalo Aguirre Beltrán s/n, Zona Universitaria, C.P. 91000 Xalapa, Veracruz, México.

#### María de Jesús Martínez-Hernández

Facultad de Ciencias Agrícolas, Circuito Gonzalo Aguirre Beltrán s/n, Zona Universitaria, C.P. 91000 Xalapa, Veracruz, México.

#### María del Carmen Ramírez-Benítez Karla Daniela Hernández-González

Facultad de Biología, Universidad Veracruzana, Circuito Gonzalo Aguirre Beltrán s/n, Zona Universitaria, C.P. 91000 Xalapa, Veracruz, México.

#### Jorge Molina-Torres

Laboratorio de Fitobioquímica, Departamento de Biotecnología y Bioquímica, CINVESTAV-IPN, Unidad Irapuato, Km 9.6 Libramiento Norte Carretera Irapuato-León, C.P. 36824 Irapuato, Guanajuato, Gto., Mex.

#### Mayvi Alvarado-Olivarez

Instituto de Neuroetologia, Universidad Veracruzana, Av. Dr. Luis Castelazo Industrial de las ánimas, Rubi Animas, C.P. 91190 Xalapa-Enríquez, Veracruz. México.

#### Vianey del Rocío Torres-Pelayo / vitorres@uv.mx

Facultad de Biología, Universidad Veracruzana, Circuito Gonzalo Aguirre Beltrán s/n, Zona Universitaria, C.P. 91000 Xalapa, Veracruz, México.

**R**ESUMEN: El fruto de zarzamora (*Rubus fruticosus*) es considerado alto en fibra, vitaminas, minerales y antioxidantes y se destacan por tener propiedades terapéuticas. Sin embargo, es importante determinar si estos antioxidantes aún se conservan en alimentos o frutas procesados; y brindar al consumidor información nutricional de este producto, el cual ha tenido una gran demanda en la región centro del Estado de Veracruz. El objetivo principal fue determinar los contenidos totales de flavonoides y antocianinas en la mermelada y licor de zarzamora artesanal; así como la cantidad de carbohidratos, lípidos y proteínas totales en estos productos, elaborados en Atecaxil, Veracruz, México. Se realizó un tamiz fitoquímico preliminar cualitativa y Cromatografía en Capa Fina de Alta Resolución (HPTLC, por sus siglas en inglés) en frutos, licor y mermelada artesana de zarzamora l. Los flavonoides y antocianinas totales se cuantificaron en la mermelada y licor artesanal de zarzamora por espectrofotometría, durante los tres años de producción. Con la prueba fitoquímica cualitativa, se encontró mayor presencia (+++) de alcaloides y flavonoides tanto en el fruto como en ambos productos artesanales. Se observó en la placa cromatográfica, bandas de color púrpura a púrpura claro, que son características de las antocianinas (Rf 0.2 a 0.3). Durante los tres años de producción la concentración de flavonoides y antocianinas fue similar, comparando entre los productos, hay mayor cantidad de flavonoides totales en la mermelada (43,6 mg) que en el licor (16,0 mg). Las antocianinas se encontraron en mayor cantidad en la mermelada de mora (59,3 mg) que

en el licor de mora (23,6 mg). El licor de zarzamora artesanal se encontró un valor energético de 82,48 kJ/19,7 kcal por 15 mL, 3,5 g de carbohidratos, 0,1 g de lípidos y 0,1 g de proteína total, 33 porciones por envase (cada porción de 15mL). En la mermelada de zarzamora artesanal se encontró un valor energético de 110.11kJ/26.3 kcal por 10 g, 6g de carbohidratos, 0.1 g de lípidos y 0.1 g de proteína total, 34 porciones cada envase (cada porción de 10 g). Estos productos artesanales podrían competir en el mercado; debido que aún se preservan los flavonoides y antocianinas, y su valor energético podría proporcionar información importante al consumidor.

Palabras claves: Alimento, productos comerciales, Rubus sp., antioxidantes, fitoquímicos.

ABSTRACT: The fruit of blackberry (Rubus fruticosus) bush is considered high in fiber, vitamins, mineral and some components are considered as antioxidants, they stand out by their therapeutic properties. Nevertheless, it is important to determine if these antioxidants are still retained in processed food; and to provide the consumer nutrimental information of this product, which has been in great demand in the central region of the State of Veracruz. The main objective was to determine the total contents of flavonoids and anthocyanins in jelly and handmade blackberry liqueur; as well as the amount of total carbohydrates, lipids and proteins contained in these products, produced in Atecaxil, Veracruz, Mexico. A preliminary phytochemical analysis and High Performance Thin Layer Chromatography (HPTLC) were performed on fruits, liquor and jelly of blackberry handmade. Total flavonoids and anthocyanins were quantified by spectrophotometry, during the three years of production. In the three samples there was a higher presence of alkaloids and flavonoids (+++). Bands purple color to light purple were observed on the chromatography plate (Rf 0.2 a 0.3), they are characteristic of anthocyanins. During the three years of production, the concentration of flavonoids of flavonoids and anthocyanins were similar, comparing between the products, there is a greater concentration of total flavonoids in jelly (43.6 mg) than in liqueur (16.0 mg). Anthocyanins were found in higher concentration in the blackberry jelly (59.3mg) than in the blackberry liqueur (23.6 mg). Both products recovered total flavonoids and anthocyanins, after the processing of the blackberry fruit. The handmade liqueur had an energetic value of 82.48kJ/19.7 kcal per 15 mL, 3.5g of carbohydrates, 0,1 g of lipids and 0.1 g of total protein, 33 serving per container (each portion or serving of 15 mL). In the artisanal blackberry jelly there was an energetic value of 110.11kJ/26.3 kcal per 10 g, 6g of carbohydrates, 0.1 g of lipids and 0.1 g of total protein, 34 portions each container (each portion of 10g). These products could compete in the handmade market, because the total flavonoids and anthocyanins are still preserved during the fabrication of the product; which could provide important information to the consumer.

Key words: Functional foods, Craft products, Rubus sp., Antioxidants, Phytochemical.

#### **I**NTRODUCTION

The fruit of blackberry belonging to the genus *Rubus* of the Rosaceae family, is a perennial shrub with aggregate fruit and is considered widely varied all over the world (Li *et al.*, 2022; Li *et al.*, 2012). In the zone of the mountain mesophilic forest of the central region of the state of Veracruz, it is one of the states in which blackberry is cultivated, as wild blackberries as introduced ones; and they are consumed mainly as fresh fruit, in jellies, and a liqueur known as "morita". These fruits are obtained from the cultivars in the field of localities of the central region of the State of Veracruz. For example, Atecaxil, Ixhuacan de los Reyes, Veracruz, Mexico is a locality rich in culture and most of the population has been devote to the sale of goat cheese, breeding creole poultry, egg production and the elaboration of handmade products derived from bush berry, as an economic sustenance source. These products are made by a local family which is dedicated to producing jelly and handmade liqueur made of blackberry (*Rubus fruticosus*). Previous research has reported that the fruit of bush berry (*Rubus ssp*) has a great number of polyphenolic components (Robinson *et al.*, 2020), and is a rich source of minerals, vitamins (such as vitamins E and C), calcium, flavonoids, anthocyanins, ellagic acid,

ellagitannins, epi/catechin, and proanthocyanidins (Baby et al., 2018). In recent years, consumers have shown increasing interest in blackberries due to their rich nutrient content, which are considered nutraceutical to human health (Li et al., 2022). The anthocyanins are natural colorants (Morata et al., 2019) and some are considered as bioactive compounds (Huang et al., 2022). They could be functional products due to presence of these phenolic compounds, which are considered as chemopreventive, antiinflammatory agents, antioxidant capacity and neuroprotective effects (Gardener et al., 2021; Zannou & Koca, 2022). Also, diets higher in flavonoids appear nutritionally beneficial in the prevention of cardiovascular disease (Parmenter et al., 2020; Santacruz Cifuentes, 2011). However, it is unknown if antioxidants are still preserved in processed products. Several studies have investigated the effects of juice processing on blueberry polyphenolics, for example, in black mulberries jelly (Morus nigra), the content of antioxidants in the processed fruit it decreases (Tomas et al., 2017), but approximately, 60-70% of the fruit anthocyanins were retained in the final juice (Tomas et al., 2015); however, these variations should not be generalized for all processed foods, because mashing and pressing some times are effective for the recovery of fruit polyphenolics into the juice fraction. Thus, it is important to evaluate the presence of these phenolic compounds in processed products, in this case, handmade products, and we could contribute to the knowledge and relevant nutrimental information to producers and consumers. The handmade products do not have nutrimental information available for consumers and have had a huge demand in the central region of the State of Veracruz. The main objectives of this study were 1) to determine the presence of flavonoids and total anthocyanins in cultivated fruits "Rubus fruticosus", as well as in derived products, liqueur and handmade blackberry jelly, and 2) to determine carbohydrates, lipids, proteins total and the energetic value measured in calories and number of portions by container of handmade blackberry products elaborated in Atecáxil, Ixhuacán de los Reyes, Veracruz, Mexico.

#### **M**ATERIALS AND METHODS

#### Collection of handmade products

The locality of Atecaxil, geographically is located in Municipality of Ixhuacan de los Reyes of the state of Veracruz, Mexico, in the coordinates Length (dec) -97.076389 and Latitude (dec) 19.373611, with a height of 1580 meters above sea level (msnm by its abbreviation in Spanish). Jars of liqueur and blackberry jelly bottled of the community of Atecaxil, Ixhuacan de Los Reyes, Veracruz, were obtained in 2017, 2018 and 2019; each product by years were kept in freezer at -20 °C with thermostat (Black&Decker®) until their analysis at laboratory. Likewise, fruits of blackberry (*Rubus fruticosus*) cultivated in the town of Atecaxil, Ixhuacan de Los Reyes, Veracruz (Figura 1), were obtained during the months of March and May of each year. These were placed in a paper bag and stored in a container to be taken in the freezer at -20 °C (Black&Decker®) until their analysis at laboratory by each year, and to avoid degradation of the phytochemicals.

#### Sample ethanolic extraction

Extracts of cultivated blackberry fruits (*Rubus fruiticosus*) and jelly (5 g) were made by sonication with 100 mL ethanol to 96% (Cole-Parmer® 8891 ultrasonic bath) for 30 minutes, until the sample was depleted. Obtained extracts were concentrated under reduced pressure in a rotary evaporator (Avante®, RE100-Pro) at a controlled temperature ( $45^{\circ}$ C). The blackberry liqueur was filtered with Whatman paper number one, and it was kept, previously labeled, in freezer at -20°C (Black&Decker®) for their analysis and to avoid degradation of the phytochemicals (Carmona-Hernández *et al.*, 2014).



Fig. 1. A) Cultivated blackberry fruits (*Rubus fruticosus*), B) Fruits collected of *Rubus fruticosus*, C) Mrs. Lupe preparing blackberry jelly in casserole.

#### Preliminary phytochemical screening

The identification of the secondary metabolites present in ethanolic extracts of fruits (*Rubus fruticosus*), jelly and blackberry liqueur, is carried out by qualitative tests for alkaloids, flavonoids, phenolic compounds, triterpenes/sterols, coumarins and saponins, using described method by (Domínguez, 1979) and modified by Carmona-Hernández *et al.*, (2014). All samples were analyzed by each year, If there was no variation in preliminary phytochemical screening, it was considered to analyze the handmade blackberry product at the end of production, only in year 2019.

#### High-Performance Thin-Layer Chromatography (HPTLC)

The ethanolic extracts of fruit, liqueur and jelly of blackberry were applied on a glass plate Silica gel Merk 60 F254 (2 µm thick) of 10 by 10 cm. 3 µL of each sample were injected onto the plate, including 4 standards (routine, quercetin, chlorogenic acid and caffeic acid), bandwidth of 8 mm, first application on the X axis to 15.8 mm, first application on the Y axis 15 mm, distance between bands of 11.4 mm, application speed 10s/µL. The plates were developed using a mobile phase of n-butanol - acetic acid - water (50:10:20: V/V), drying of the plate for 30s, followed by a saturation of 20 min at room temperature; and after the development a drying of 10 min. The derivatization was performed with Anisaldehyde-sulfuric acid (a universal reagent for natural products) by heating the plate at 100°C for 10 min (Agatonovic-Kustrin *et al.*, 2019; Wagner & Bladt, 1996). The derivatization was used for target-directed identification of biologically active molecules separated on chromatographic plates. The chromatographic plates were developed and visualized in visible light and at wavelengths at 254 and 366nm with the CAMAG AUTOMATIC TC SAMPLER 4 (CINVESTAV-IPN, Irapuato Unit, Guanajuato).

#### **Total flavonoids content**

The colorimetric method of aluminum chloride was used to determine total flavonoids (Martínez-Cruz *et al.*, 2011; Zhishen *et al.*, 1999). 200 $\mu$ L of fruit extracts, jelly and blackberry liqueur were taken and mixed with 800 $\mu$ L at 96% ethanol, plus 1mL of aluminum chloride at 2%. Subsequently, the mixture was incubated for 30 min at room temperature and in the absence of light. The absorbance was measured at 510nm in a spectrophotometer (JENWAY-GENOVA<sup>®</sup>). For the calibration curve, Quercetin solutions between 0-50 $\mu$ g were used (Sigma Aldrich®). Total flavonoid content was expressed as milligrams (mg) Quercetin (EQ) equivalents/100 g of sample. This procedure was carried out with each of the extracts obtained in triplicate.

#### Total anthocyanins content

Determination of total anthocyanins was carried out by the method described by (Di Stefano & Flamini, 2008) and method modified by (Ivanova *et al.*, 2010). 200 $\mu$ L of fruit sample, jelly and blackberry liqueur were mixed with 1800 $\mu$ L of an ethanol solution/ H<sub>2</sub>O/HCl (70:30:1), pH 2 and immediately afterwards the absorbance at 540 nm was measured in a spectrophotometer (JENWAY-GENOVA<sup>®</sup>). The content of total anthocyanins was calculated using the following equation:

AT 540nm (mg/L) = A540nm\*16.7\*d

Where:  $A_{540}$ nm is the absorbance and, *d* is the dilution and 16.7, molar extinction coefficient of malvidin-3-glucoside, using described method modified by Ivanova *et al.*, (2010).

#### **Statistical Analysis**

Total flavonoids and anthocyanins content were reported as means  $\pm$  standard deviation of five samples by years. The data were subjected to analysis of variance (ANOVA) to repeated samples and were compared with a Tukey test (P  $\leq$  0.05). All statistical analyses were performed using the STATISTICA® program for Windows.

#### **Basic Nutrimental Profile of Products**

Random containers were obtained of each product (10 containers), blackberry liqueur and jelly. To quantify carbohydrates, lipids and total proteins, from each sample of blackberry liqueur and jelly. First were obtained the constant weight to determine dry weight and percentage of humidity in the samples to be analyzed. Then, 5g of jelly and 5mL of liqueur were used to determine the amount of carbohydrates, lipids and total proteins following the protocol (Analytical Official Chemists Association guide was considered, "AQAO by its abbreviation in Spanish" 2015).

#### **Total Carbohydrates Quantification**

It was used the method described by (López-Legarda *et al.*, 2017) which consisted of taking aliquots of 2mg of blackberry liqueur and blackberry jelly in which 1mL of phenol were added at 5% and 5mL of H<sub>2</sub>SO<sub>4</sub> absolute concentration. The tubes were shaken for 30 seconds approximately, until to be mixed. The reaction of samples was stopped in cold bath during a period of 20 minutes, once the reaction was stopped, a reading in the spectrophotometer (JENWAY-GENOVA<sup>®</sup>) was carried out at 490nm. The values obtained of absorbance were helpful to plot a graph which is very crucial in estimating carbohydrate content. The calibration curve took place with a glucose standard (100mg/mL), repeating the same method used previously.

#### **Total Lipids Quantification**

It was used the method modified by (Lykke & Padonou, 2019), to determine the total Lipids Quantification, which consisted of homogenizing 5g of liqueur dry extract (extracts previously obtained to eliminate alcohol, humid and impurities) with 10mL of chloroform and 20mL of methanol; the same procedure was used with 5g of blackberry jelly (extracts previously obtained). Later, another 10mL of chloroform were added and homogenized for 1min. When the procedure ended, 10mL of water were added and it was homogenized for 1 more min. The mixture was filtered with Whatman paper number 1, the paper was set in a Buchner funnel in vacuum. The filtered mixture was transferred to a test tube of 50mL, once again the filter was washed with 10mL of chloroform and methanolic phase, since separation was obtained, methanolic superior phase was withdrawn with a Pasteur pipette and the volume of chloroform phase was transferred to a flask previously weighed (first weighting of flask). Later, it was set in water bath to 50°C in order to evaporate chloroform phase and the remaining was dried over anhydrophosphoric in a desiccator in vacuum. After that, the flask was weighed for a second time, and 5mL of chloroform was added to extract lipids. This solution was poured into another

flask, cleaned and dried previously. The flask containing lipids evaporated one more time and it was weighed for third time.

The next calculation was carried out.

Lipids weight = weight of flask that containing lipids evaporated X total volume of chloroform used (30mL) / evaporated chloroform volume.

#### **Total Protein Quantification**

It was used the Biuret method (Ceballos Luna *et al.*, 2018; Coutiño *et al.*, 2015), which consisted of adding 0.5mL of liqueur and 0.5g of blackberry jelly in test tubes. In each test tube containing the samples, 1mL of NaCl were added in 0.9% and 3mL of Biuret reactive, subsequently, it was mixed till homogenize. The obtained mixtures were heated to 50°C in water bath for 10min, when it finished a reading was carried out in the spectrophotometer at 540nm a calibration curve was taken in albumin standard (100mg/mL), repeating the procedure used before.

#### **Elaborating Nutritional Information**

To elaborate the nutritional information of homemade products of berry and to obtain the energy calculation from conversion factors, it was used the method described by (Salvador Badui Dergal, 2006) and (Pérez Grana, 2013) while consisted in used accuracy of food composition tables in the determination of nutrients. As well, it was determined the amount of carbohydrates, lipids and total proteins by each portion of blackberry liqueur and jelly, 15mL of liqueur and 10g of homemade blackberry jelly. The reported values were converted into the same units and synthetized in a table. The Mexican official standard NOM-051-SCFI-1995 was used to reference the table of homemade products.

#### **R**ESULTS AND **D**ISCUSSION

#### Preliminary phytochemical analysis

The qualitative analysis for secondary metabolites in the fruit of blackberry, jelly and handmade liqueur in three years showed a similar pattern. Mainly displayed presence of alkaloids and flavonoids; and minor presence of saponins, terpenes and steroids. However, in all samples, coumarins had a low presence (Table 1, only showed results in 2019). These results were similar during the three years and are therefore an important parameter to evaluate the phenolic contents in Rubus fruticosus of Atecaxil, Ver.; mainly the anthocyanins and total flavonoids content in fruit, jelly and handmade blackberry liqueur was carried in the final year of production on 2019. (Mulero et al., 2011) reported antioxidant activity in all 3 types of wine elaborated and found no differences in the concentrations of the different types of phenolic compounds in wine made with the 3 different methods, and during 3 months of storage showed a similar pattern. Thus, we still found the prevalence of alkaloids and flavonoids in the fruit of blackberry (Rubus fruticosus) and in processed blackberry products of Atecaxil, Ver. These qualitative findings, depends, not only on the quantity of sample, but also on the bond and/or interaction of these compounds with other molecules, on the location in the food matrix, and the presence of other bioactive compounds into samples (Minatel et al., 2017). During the cooking process, the temperature thermal may to destruction of cell wall and other subcellular components, stimulating the release of these compounds (alkaloids, flavonoids, saponins, terpenes and steroids) and were stable during the manufacturing process. Several research studies have shown increase in the phenolic compound levels, as well as in the antioxidant activity after cooking (Leong & Oey, 2012; Murador et al., 2016). It's important pointing, there are indications that the retention of phytochemicals and the antioxidant properties are present after the cooking and vary considerably between the different vegetables and methods used in their preparation ((Jiménez-Monreal et al., 2009; Minatel et al., 2017; Pellegrini et al., 2009). However, is important to remember that the exact composition of blackberries is highly dependent on the cultivars, cultural practices and numerous preharvest factors, especially

climatic and soil conditions (Vergara et al., 2016). Mainly the flavonoids are a group of natural substances with variable phenolic structures and are widely distributed in plants; recently, 26 phenolics compounds were identified and quercetin and isoquercitrin were the predominant phenolic compounds in the fruit in Rubus ulmifolius (Schulz et al., 2019). In year previous, have been reported thousands of phenolic antioxidant compounds exist and are classified into several categories based on structural similarities (Craft et al., 2012), in the maturation stages (Schulz et al., 2019). For example, separated phenolics into seven categories: phenolic acids, coumarins, flavonoids, isoflavonoids, stilbenes, lignans, and phenolic polymers (tannins), hydroxybenzoic acids, flavan-3-ols and anthocyanin. These groups differ from each other in functional group placement, or the addition of a chemical moiety as in glycosylation. These results reinforce the prevalence potential of flavonoids alkaloids, saponins, terpenes and steroids in blackberry fruit (Rubus fruticosus) and liqueur and handmade jelly from Atecaxil, Ixhuacan de Los Reyes, Veracruz (Table 1). On the other hand, there are limited information is available on how different processing methods in processed products, impact on the polyphenolic content of the fruit (Brownmiller et al., 2009), which is implied by an increase of its bioavailability in the human body when they consume it.

 Table 1. Qualitative result of preliminary phytochemical analysis of samples from blackberry fruit, liqueur and handmade jelly from Atecaxil, Ixhuacan de Los Reyes, Veracruz, on year 2019.

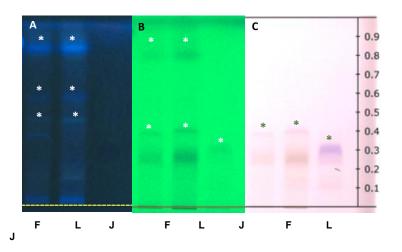
Compound	Alkaloids		Flavonoids		Saponins		Coumarins	Terpenes/ steroids		
Products	Qualitative Tests									
Troutes	М	D	W	А	С	S	F	SL	Со	LB
Blackberry fruit	++	+++	++	+	+++	+++	++	n/d	+	+
Jelly	-	++	++	++	+++	++	-	n/d	+	-
Liqueur	-	++	+++	-	+++	++	++	n/d	+	++

Alkaloids: M= Mayer Test, D= Drangendorff y Test, W= Wagner Test. Flavonoids: A= H<sub>2</sub>SO<sub>4</sub> Test, C= FeCl<sub>2</sub> Test, S= Shinoda Test. Saponins: F= Foam Test, SL= Lieberman Test. Coumarins: Co= Fluorescence Test. Triterpenes and/or Sterols: LB= Lieberman-Bouchard Test. Intensity: major +++, medium ++, low +, null – and n/d non-determined.

#### High-Performance Thin-Layer Chromatography

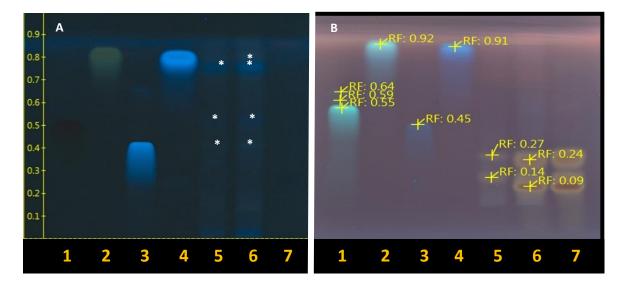
The plate observed in visible light (RT-White), was found in the sample of fruit and liqueur of blackberry a band of brown to red light brick color with a Rf of retention 0.25 and another thin band with a Rf 0.4. While, in the jelly sample, a band from purple to light purple was observed with a Rf 0.3. Also, a wavelength of 254 nm, light bands were observed in the three samples, with a Rf 0.25, they were observed in visible light; but with a single variation of bands observed with a Rf 0.8 (Fig. 2). Finally, at a wavelength of 366nm, bands with a similar Rf 0.4, 0.6 and 0.85 were observed both in fruit and blackberry liqueur. On the other hand, a Rf 0.15 and 0.45 were observed in liqueur (Fig. 2). These similarities and differences indicate that there are compounds shared, at least, some similar chemical structures or they are similar compounds. Observed coloration in visible light between fruit, liqueur and blackberry jelly, can indicate that both in fruit and products, there are compounds that are maintained after the elaboration process, bottled and shelf life. Also, the pigments continue to be observed. Similar studies in blackberry fruits of the species Rubus adenotrichus Schltdl and other wild blackberries (Rubus ssp) from the central region of Veracruz, have reported presence of these pigments (Aguilera-Otíz et al., 2011; Morata et al., 2019), known as anthocyanins (Martínez-Cruz et al., 2011). Aguilera-Otíz et al., (2011) reported that intensification of blue coloration is due to increase of phenolic ring hydroxyls, while introduction of methoxyls cause the formation

of red color. Also, (Morata et al., 2019) reported that the color of natural anthocyanins covers most of the visual spectra from yellow - orange to blue-violet. The stability of anthocyanins depends on several parameters, such as pH, temperature, and oxidative conditions, but they are normally quite stable in acidic media. Likewise, it has been reported that in aqueous solutions, pigments are basically stable, and they have an intense red color, and at pH above 7, quinoidal shapes are present (A, A-) purple color that is degraded rapidly because of oxidation with air; sometimes when oxygen is excluded from the system, color deterioration is not observed (Garzón, 2008; Morata et al., 2019). Thus, during the elaboration and conservation of handmade products from the locality Atecaxil; these chemical factors mentioned before, had not effect on pigments, since purple color or light purple or blue and brown tones to light brick red are still observed in both products; aversely, temperature applied in the elaboration of blackberry jelly did not affect, and the cane alcohol added to the blackberry liqueur, could detonate particular features in each product. Therefore, during the elaboration process of these products, it is possible to detect pigments light brick brown-reddish coloration in blackberry handmade liqueur, meanwhile in blackberry jelly from purple to light purple color can be observed (Fig. 2-C and 3). In this study, we found pigments still when submitted to thermal processing, in water or ethanol (Fig. 2). (Kalt et al., 2020), reported two blueberry polyphenolic compounds, including both flavonoid and nonflavonoid types. There are abundant nonflavonoid polyphenolic compounds in blueberries are the hydroxycinnamic acid esters (especially chlorogenic acid). In our results, we found bands con Rf similarities to standard chlorogenic acid (Fig. 3 A).



**Fig. 2.** High performance thin layer chromatography of Fruit (F), Liqueur (L) and blackberry Jelly (J) observed at wavelengths of 366nm (A), 254 nm (B) and RT White (C); observing the separation of compounds reflected in bands, with similarity and difference between Rf's each sample [Indicated with an asterisk in white (A and B) and green (C)].

It can be observed, bands Rf similar to the standards used (Rutin, Quercetin, Chlorogenic Acid and Caffeic Acid) whit the technique not derivatized (Fig. 3-A), between fruits and liqueur showed similar results. While that derivatized with Anisaldehyde-Sulfuric Acid (AS) reagent, it is reflected the separation of others compounds in Rf's each sample (Fig. 3-B), i.e., all samples revealed additional bands in the upper part of the chromatograms, showed similar results (Rf 0.09 to 0.27) between fruits, liqueur and handmade blackberry Jelly. Among the factors that affect the leaching of matrix compounds, we can include the polarity of medium used; such as water, allow changes in the phenolic compound levels. In contrast, if the medium is nonpolar (use of oil for frying, in both deep frying and pan frying), the loss of compounds is lower due to the lack of diffusion or migration to the medium or, may be due to the amount of fruit used to make the homemade jelly and the fruit, flavonoids were retained more in the jelly that liqueur. Blackberry contain abundant phenolic compounds, including anthocyanins (Cyanidin Delphinidin, Malvidin, Peonidin), flavonols and chlorogenic acids (Sandoval et al., 2020). Some of these compounds are pigments that impart pleasant and characteristic colours to the fruits. Another hand, (Li et al., 2022) found phenolic acids such as p-hydroxybenzoic acid, chlorogenic acid, coumarin, and syringic acid are the most widely available polyphenols in fruit juices (samples of blueberry, cherry and mulberry). The Rutin in mulberry juice is the highest polyphenol (420.87  $\mu$ g/g). Therefore, the composition and content of phenolic substances are different in fruit juice. This authors, descripted in previous research that polyphenolic compounds are the material basis for the flavor, color, and nutritional properties of fruits, which directly affect the taste and quality of fruits and fruit-based processed foods (Maragò et al., 2016), or depending on the type of used solvent, it can interfere in the number of compounds present in fruits (Lee et al., 2012; Li et al., 2022). While Minatel et al., (2017) mentioned that flavonols improved the color stability of mulberry juice during storage. In addition, blueberries are one of the richest sources of anthocyanins among common fruits (Wu et al., 2006) and anthocyanins are the pigments that confer red, blue, and purple coloration. During berry ripening, anthocyanin content rises dramatically to provide a visual cue to distinguish between early to fully ripe fruit (Kalt et al., 2020). Finally, in the three samples, indicate that there is presence of flavonoids and anthocyanins in blackberry jelly and liqueur (Fig. 3).



**Fig. 3.** High performance thin layer chromatography of Fruit (F), Liqueur (L) and blackberry Jelly (J) observed at wavelengths of 366nm. A) Not derivatized and B) Derivatized with Anisaldehyde-Sulfuric Acid (AS) reagent. 1: Rutin, 2: Quercetin, 3: chlorogenic acid; 4: Caffeic Acid, 5: blackberry fruit, 6: blackberry liqueur, 7: blackberry jelly. It is observing the separation of compounds reflected in bands, with similarity and difference between Rf's each sample [Indicated with an asterisk in white (A)].

In the table 2, showed contend similar in total flavonoids and anthocyanins in fruit samples, handmade blackberry jam and liqueur, do not show significant year-by-year differences (Table 2). When comparing anthocyanin concentrations between handmade products, there are a higher concentration of anthocyanins in blackberry jelly and lower concentration in liqueur and fruit (Table 2), the fruit anthocyanins were retained in the final jelly, the temperature did not affect the presence of anthocyanins. The anthocyanins quantity found in handmade products may be due, firstly, to the amount of fruit used to make both products and state of maturity of the fruit as has been reported in Zarzamora (*Rubus ulmifolius*) by Borrego Corchado, (2018).

Similarly, previous studies by (Li et al., 2022) reported an increment of anthocyanin content whit the increased gradual of the ripening stages of fruits. Also, indicated anthocyanins content in blackberry does not decrease with respect to the thermal processing of fruit, so they propose the idea that they are preserved and can be a good source of antioxidants (Bernal-Roa, 2012; Garzón, 2008). There is evidence that in the blackberry pulp have a concentration of monomeric anthocyanins between 6.05 to 7.37 mg/L (Rodríguez-Pérez et al., 2010) or ranged from 70.3 to 201 mg/100 g FW (fresh weight). In other studies have reported an average of 12.3 mg of anthocyanins (equivalent of malvidin-3-glucoside) for each gram of dried fruit of Rubus adenotrichus species (Martínez-Cruz et al., 2011). Previous reports indicated that blueberries sample is quite different from those published for blueberries from other locations, being the major anthocyanins found peonidin-3-O-arabinoside and delphinidin-3-O-arabinoside (37.43  $\pm$ 4.76 and  $34.43 \pm 3.28$  mg/100 g fresh weight, respectively) followed by malvidin-3-Oglucoside and petunidin-3-O-rutinoside (Johnson et al., 2020; Yousef et al., 2013). Mustafa et al., (2022) also found anthocyanins were the main phenolic constituents in blueberry and strawberry. Furthermore, the higher total phenolic content in the blueberry fruit and jam justified their greater antioxidant capacity measured by DPPH free radical assay, compared to strawberry. Elez Garofulić et al., (2012) reported that anthocyanins are the predominant wine pigments, transferred to wine from both fruit skin and pulp during the maceration process and the concentration oscillated in an extremely wide range, from 5.07 mg/L (CBW 2) to 217 mg/L (CBW 7), but there is difference between the conventional wines (76.2 mg/L) and organic wines, was considerably lower (53.5 mg/L) (Amidžić Klarić et al., 2020). This is in accordance with the study of Johnson & Gonzalez de Mejia, (2012), that reported concentrations of total anthocyanins in blackberry wines in the range from 10.71 mg/L to 191.95 mg/L (expressed as cvaniding-3-glucoside equivalents) and an average concentration of 75.56 mg/L. Mudnic *et al.*. (2010) also reported an extremely wide range of total anthocyanin values for blackberry wines, i.e., from  $13.4 \pm 3$  to  $164 \pm 3$  mg/L (expressed as malvidin 3-glucoside equivalents) and found it to be comparable to total anthocyanins in red grape wine. In contrast, the results of the present study did not indicate a significant difference in total anthocyanins between the years, but there is significant difference between products. These variations may be due to studied species and the amount of used sample or location, cultivar, and time of harvest; as is reported by other authors (Bunea et al., 2011; Fia et al., 2018). However, the anthocyanin, cyanidin-3-Oglucoside (C3G) are consistently the predominant phenolic antioxidant found in blackberries and have been shown to have prominent bioactivity (Schulz et al., 2019). Among the factors that affect the prevalence of compounds, is the polarity of medium used. Polar mediums, such as water and ethanol, allow changes in the phenolic compound levels (Gardener *et al.*, 2021; Pellegrini et al., 2009).

Sample	]	Fotal Flavono	ids	Total Anthocyanins			
	2017	2018	2019	2017	2018	2019	
Blackberry fruit	$20.5\pm2.8$	26.1 ± 3.6	$24.3 \pm 1.0$	$12.37\pm1.8$	$16.19\pm0.7$	$13.26\pm5.7$	
Jelly	$38.8\pm 6.8$	$43.6\pm1.3$	$38.0\pm4.06$	$59.3\pm4.4$	$54.6\pm3.8$	$52.1\pm2.6$	
Liqueur	$16.0 \pm 3.6$	$24.0\pm3.4$	$20.0 \pm 1.7$	$20.5\pm3.8$	$23.6\pm0.2$	$22.1 \pm 3.2$	

**Table 2.** Total flavonoids and anthocyanins in fruit samples, handmade blackberry jelly and liqueur from Atecaxil,Ixhuacan de Los Reyes, Veracruz, during three years of production 2017 to 2019.

*Values are represented in miligramos (mg)and standard deviation from 3 replicates by years (ANOVA-Tukey test, P* $\leq$ 0.5).

Total Flavonoids expressed in milligram (mg) EQ/100g

Total Anthocyanins expressed in milligram (mg) of malvidin-3-glucoside/100g

On the other hand, when comparing total flavonoids concentrations between handmade products, there is a higher concentration in blackberry jelly and lower concentration in liqueur and fruit (Table 2). In blackberry jelly there is more fruits incorporated, while in blackberry liqueur has less fruit used for its preparation, according to the comments of the producing family. Similarly, other authors mention that black mulberry juice is rich in phenolic acids compared to the fruit and anthocyanins are highly retained during juice processing, approximately 60-70% (Tomas et al., 2015, 2017), effectively, recover polyphenolics into the juice. Other authors have found (+)-Catechin and its isomer (-)-epicatechin, are consistently the most abundant flavan-3-ols (class of flavonoids) in blackberries (Robinson et al., 2020). (Li et al., 2022) analyzed polyphenols in fruit juice samples of blueberry, cherry and mulberry, they found that the yields of polyphenols extracted by ethanol from fruit juice samples were obviously higher than those by water and acetone. They determined how canning, puréeing and juicing of blueberries, as well as storage of processed products at 25°C, influenced the retention of chlorogenic acid, total flavonols, total anthocyanins, and total procyanidins. The retention of flavonols (57-99%) and chlorogenic acid (64-100%) was greater than that of anthocyanins (42-72%) and procyanidins (19-78%). In non-clarified juices retained higher levels of chlorogenic acid, total flavonols, and total anthocyanins than clarified juices, but clarified juices contained higher levels of total procyanidins. The 97% retention of chlorogenic acid in non-clarified juices was much higher than the previously reported value (53%) for non-clarified blueberry juices (Skrede et al., 2000). It was also found in organic blackberry wines (BW) than the concentration of caffeic acid and p-coumaric acid, both being higher in the organic BW samples than in conventional vines. While than the concentration of total flavonoids in the analyzed samples of blackberry wine ranged from 161 to 774 mg/L, and the mean concentration between the organic and conventional group of samples did not differ (Amidžić Klarić et al., 2020). Another factor that affects the prevalence of compounds is the polarity of medium used, such as water and ethanol, allow changes in the phenolic compound levels (Gardener et al., 2021; Pellegrini et al., 2009). In any case, processing the fruit facilitated its subsequent extraction in our studies, we are recovering total flavonoids and anthocyanins after liqueur and jelly processing of the blackberry fruit (see Table 2). In this study, we confirmed that it is conserve the total flavonoids and anthocyanins and the elaboration process does not affect the presence of flavonoids; the antioxidant properties have been associated with their phenolic composition and particularly with the high content in anthocyanins and flavonoids. Genova et al., (2016), mentioned that an appropriate management of fruit harvesting date, postharvest and processing may lead to an improvement in nutraceutical quality of juices or another product. The phytochemical quantity retained in fruits and vegetables, after the processing, depends on the stability of these compounds during the different food preparations. Molecular modifications induced by processing and the transformations that occur before the consumption are mainly related to the sensibility of the compounds to oxidation and/or isomerization (Leong & Oey, 2012). Theses researchers corroborating our results phytochemical, total flavonoids were maintained in fruit samples, handmade blackberry jam and liqueur From Atecaxil, Ver. Mex. Generally, it has been reported that blackberry (red fruits) are primary sources of flavonoids, and they can be associated with antioxidant activity (Bunea *et al.*, 2011; Geleijnse & Hollman, 2008; Lillo *et al.*, 2016; Martínez-Cruz *et al.*, 2011). The antioxidant content of blackberries and in particular their phenolic content ought to be considered as an important trait for breeding programmes (Clark *et al.*, 2011; Milošević *et al.*, 2012), as well as rural or urban areas. Minatel *et al.*, (2017) reported that even though there are innumerous studies comparing the biological actions and *in vitro* antioxidant activity of phenolics, and the function of its content in vegetables and consequently in human, there is no consensus about the best way of preparing/consuming fruits and vegetables intending preservation or to increase their antioxidant activity.

#### Total Carbohydrates, Lipids and Protein Quantification

Total carbohydrates concentration in homemade liqueur of blackberry was of  $0.84 \pm 0.5$  mg/mL and in blackberry jelly was of  $0.74 \pm 0.5$  mg/mL, studies carried out in raspberry wine (Autumn *bliss var*), have found a concentration of carbohydrates of  $1.8 \pm 0.3$  mg/mL and, in red fruit wine,  $2.2 \pm 0.47$  mg/mL (Sánchez Trujillo, 2013). Total lipids concentration in blackberry liqueur and jelly were of  $0.0009 \pm 0.2 \ \mu g/mL$  (9 mg/mL) and  $0.0013 \pm 0.15 \ \mu g/mL$  (1.3 mg/mL), respectively. This variation in quantities can be due to two important factors, the amount of fruit used or the extraction only of the fruit juice during the manufacturing of blackberry liqueur, because, in the jelly was used all fruit. For instance, during the elaboration of blackberry jelly, producers used all drupe of the fruit, including seeds, meanwhile in blackberry liqueur, it was used only juice of fruit, therefore, it is reflected a higher amount of total lipids in jelly than in juice. Preceding studies such as, Cerón et al., (2012) and Pantoja-Chamorro et al., (2017) reported that content of fatty acids presents in the fruit of castle blackberry (Rubus glaucus), they have bigger numbers of unsaturated fatty acids within blackberry pips, such as, linoleic acid (61.6%) and linolenic acid (25%), attributing beneficial properties to human being. However, Zafra Rojas, (2019) reported low presence of lipids in subproducts of Rubus fruticosus. For example, in Zarzamora sauces (Rubus fruticosus), a low amount of fat was found for each sample of 100 gr (Ceballos Luna et al., 2018). Lastly, the amount of proteins present in blackberry liqueur was  $0.001 \pm 0.047$  mg/mL and in blackberry jelly,  $0.004 \pm 0.031$ mg/mL of proteins were found. The seeds, fruits and leaves are important sources for both protein and fat. Fruits generally have lower content of protein, fat and minerals, but these components are still present. Studies done in red wine, found 0.001 mg/mL of proteins, while in blackberry jelly and strawberry jelly from well-known brands report in the label 0.002 mg/mL and 0.003 mg/mL of proteins, respectively (De Rosso et al., 2009; Kassara et al., 2022). In blackberry sauces, proteins totals ranged from 7 to 9% for each sample of 100 gr (Ceballos Luna et al., 2018).

In blackberry liqueur had an energetic value of 18.9 kcal/15 mL, having in total 33 portions each container, meanwhile, in blackberry jelly, it was found an energetic value of 27.1 kcal/10 g, having around 34 portions by container (Table 3). Information on the nutrient composition of food is essential to estimate adequate nutrient intake both at individual and group levels (Joyanes & Lema, 2006). Vergara *et al.*, (2016) described that blackberry plays an important role in human nutrition, due to the elevated content of certain bioactive compounds including ascorbate, anthocyanins, phenolic acids, carbohydrates and proteins. In this present research, we corroborating that the handmade products could be highly competitive in the market. Likewise, there is not a significant difference when we comparing the two handmade products in the three years (data not relevant), while we can explain the color and unique flavor of the products. In addition, we provide the consumer, the amount of macronutrients ingested in each serving of both handmade products from Atecaxil, Ver., Mex, important knowledge for your health.

Table 3. Total macronutrients in handmade products of blackberry (Rubus fruticosus) from Atecaxil, Veracruz,
Mexico in years 2019.

Blackberry Liqueur	Blackberry jelly
33 servings per-container/652.6 kcal	34 servings per-container/894.3 kcal
Serving Size 1 cup (15 mL)	Serving Size 1 tablespoon (10 g)
Energetic value 82.48kJ/19.7 kcal per 15 mL	Energetic value 110.11kJ/26.3 kcal per 10 g
Total Carbohydrate $3.5 \pm 0.48$ g	Total Carbohydrate $6 \pm 0.51$ g
Lipids 0.1 ± 0.2 g	Lipids $0.1 \pm 0.15$ g
Protein $0.1 \pm 0.05$ g	Protein $0.1 \pm 0.039$ g

#### CONCLUSIONS

In handmade blackberry products, there are presence qualitative of flavonoids and alkaloids; and minor presence of saponins, terpenes and steroids. In both products, is possible to observe pigments of purple color or light purple or blue and brown tones to light brick red. This color is plant pigments, and the anthocyanins are biologically active, water soluble and are responsible for blue, purple, and red colors, especially in fruits and blooms. Therefore, handmade blackberry products of Atecaxil, Ixhuacan de los Reyes, Veracruz., continued preserving the flavonoids and anthocyanins after of its manufacture, there was no difference in the flavonoid and anthocyanin contains between years, but there was difference in type products. These products could have antioxidant potential, as has been reported in blackberry fruit by other authors. Furthermore, we provide to consumer relevant information about metabolites source, flavonoids and total anthocyanins, which are important in the food field for human beings. Total macronutrients in handmade Blackberry Liqueur and jelly found, are relevant to new sources of information about total carbohydrate, lipids and protein for the Consumers.

#### AACKNOWLEDGMENT

We thank the student Álvaro Rafael Ruiz Rodríguez for being part of the work team and for having contributed part of the experiments of this work during his professional training and studies. To the family that produces liquor and handmade blackberry jelly from Atecaxil, Ixhuacan de los Reyes, Ver, Mex., for their hospitality and acquisition of the handmade products. SIVU Project, Faculty of Biology, UV. Xalapa, Ver., Mex.

#### LITERATURE CITED

- Agatonovic-Kustrin, S., Kustrin, E., Gegechkori, V., & Morton, D. (2019). High-Performance Thin-Layer Chromatography Hyphenated with Microchemical and Biochemical Derivatizations in Bioactivity Profiling of Marine Species. *Marine Drugs*, 17(3), 148. https://doi.org/10.3390/md17030148
- Aguilera-Otíz, M., Reza-Vargas, M. del C., Chew-Madinaveita, R. G., & Meza-Velázquez, J. A. (2011). Propiedades funcionales de las antocianinas. *BIOtecnia*, 13(2), 16–22. https://doi.org/10.18633/bt.v13i2.81
- Amidžić Klarić, D., Klarić, I., Mornar, A., Velić, N., & Velić, D. (2020). Assessment of Bioactive Phenolic Compounds and Antioxidant Activity of Blackberry Wines. *Foods*, 9(11), 1623. https://doi.org/10.3390/foods9111623

- Baby, B., Antony, P., & Vijayan, R. (2018). Antioxidant and anticancer properties of berries. *Critical Reviews in Food Science and Nutrition*, 58(15), 2491–2507. https://doi.org/10.1080/10408398.2017.1329198
- Bernal Roa, L. J. (2012). Evaluación de las Propiedades Bioactivas de Mora (*Rubus glaucus*) y Agraz (*Vaccinium meridionale Swartz*), en Fresco y Durante Procesos de Transformación [Trabajo de grado - Maestría, Universidad Nacional de Colombia]. https://repositorio.unal.edu.co/handle/unal/11023
- Borrego Corchado, C. (2018). Evolución de compuestos de interés biológico en moras a lo largo de la maduración del fruto [Máster en Agroalimentación, Universidad de Cádiz]. file:///C:/Users/torre/Downloads/TFM%20Definitivo%20(1).pdf
- Brownmiller, C., Howard, L. R., & Prior, R. L. (2009). Processing and storage effects on blueberry (*Vaccinium corymbosum* L.) polyphenolics. *Acta Horticulturae*, 841, 347– 354. https://doi.org/10.17660/ActaHortic.2009.841.43
- Bunea, A., Rugina, O. D., Pintea, A. M., Sconta, Z., Bunea, C. I., & Socaciu, C. (2011). Comparative Polyphenolic Content and Antioxidant Activities of Some Wild and Cultivated Blueberries from Romania. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 39(2), 70–76. https://doi.org/10.15835/nbha3926265
- Carmona-Hernández, Ó., Fernández, M. del S., Palmeros-Sánchez, B., & & Lozada-García, J. A. (2014). Actividad insecticida de extractos etanólicos foliares de nueve piperáceas (*Piper* spp.) EN Drosophila melanogaster. *Revista Internacional de Contaminación Ambiental*, 30, 67–73. https://www.redalyc.org/pdf/370/37033725008.pdf
- Ceballos Luna, O., Morales Sotelo, R., López Márquez, A. A., Grapain López, O., García Gutiérrez, E., Sedeño Mota, J. C., Álvarez Campos, E., & Lozada García, J. A. (2018).
  Salsa a base de zarzamora (*Rubus fruticosus*) como potencial alimento funcional. *Revista Biológico Agropecuaria Tuxpan*, 6(2.Especial), 72–77. https://doi.org/10.47808/revistabioagro.v6i2.Especial.259
- Cerón, A. F., Osorio, M. O., & Hurtado B, A. (2012). Identificación de ácidos grasos contenidos en los aceites extraídos a partir de semillas de tres diferentes especies de frutas. *Acta Agronómica*, *61*(2), 126–132. https://www.redalyc.org/pdf/1699/169925874008.pdf
- Clark, D. J., Jørgensen, F., & Mathisen, T. A. (2011). Relationships between fares, trip length and market competition. *Transportation Research Part A: Policy and Practice*, 45(7), 611–624. https://doi.org/10.1016/j.tra.2011.03.012
- Coutiño, R., Fernández, S., & Palmeros, B. (2015). *Manual de la experiencia educativa Biología Celular* (1st ed., Vol. 1). Universidad Veracruzana. https://doi.org/10.25009/uv.2018.37
- Craft, B. D., Kerrihard, A. L., Amarowicz, R., & Pegg, R. B. (2012). Phenol-based antioxidants and the in vitro methods used for their assessment. *Comprehensive Reviews in Food Science and Food Safety*, *11*(2), 148–173. https://doi.org/10.1111/j.1541-4337.2011.00173.x
- De Rosso, M., Panighel, A., Dalla Vedova, A., Stella, L., & Flamini, R. (2009). Changes in Chemical Composition of a Red Wine Aged in Acacia, Cherry, Chestnut, Mulberry, and Oak Wood Barrels. *Journal of Agricultural and Food Chemistry*, 57(5), 1915– 1920. https://doi.org/10.1021/jf803161r
- Di Stefano, R., & Flamini, R. (2008). High Performance Liquid Chromatography Analysis of Grape and Wine Polyphenols. In Flamini Riccardo (Ed.), *Hyphenated Techniques in Grape and Wine Chemistry* (John Wiley & Sons, Vol. 1, pp. 33–79). John Wiley & Sons, Ltd. https://doi.org/10.1002/9780470754320.ch2
- Domínguez, X. A. (1979). Métodos de investigación fitoquímica (1st ed.).
- Elez Garofulić, I., Kovačević Ganić, K., Galić, I., Dragović-Uzelac, V., & Savić, Z. (2012). The influence of processing on phyico-chemical parameters, phenolics, antioxidant capacity and sensory attributes of elderberry (*Sambucus nigra* L.) fruit wine. *Croatian Journal of Food Technology, Biotechnology and Nutrition*, 7, 9–13. https://hrcak.srce.hr/file/123081

- Fia, G., Gori, C., Bucalossi, G., Borghini, F., & Zanoni, B. (2018). A Naturally Occurring Antioxidant Complex from Unripe Grapes: The Case of Sangiovese (v. *Vitis vinifera*). *Antioxidants*, 7(2), 27. https://doi.org/10.3390/antiox7020027
- Gardener, S. L., Rainey-Smith, S. R., Weinborn, M., Bondonno, C. P., & Martins, R. N. (2021). Intake of Products Containing Anthocyanins, Flavanols, and Flavanones, and Cognitive Function: A Narrative Review. *Frontiers in Aging Neuroscience*, 13, 1–14. https://doi.org/10.3389/fnagi.2021.640381
- Garzón Gloria Astrid. (2008). Las antocianinas como colorantes naturales y compuestos bioactivos: Revisión. *Acta Biológica Colombiana*, *13*(3), 27–36. http://www.scielo.org.co/scielo.php?script=sci\_arttext&pid=S0120-548X2008000300002&lng=en&tlng=es.
- Geleijnse, J. M., & Hollman, P. C. (2008). Flavonoids and cardiovascular health: which compounds, what mechanisms? *The American Journal of Clinical Nutrition*, 88(1), 12–13. https://doi.org/10.1093/ajcn/88.1.12
- Genova, G., Tosetti, R., & Tonutti, P. (2016). Berry ripening, pre-processing and thermal treatments affect the phenolic composition and antioxidant capacity of grape (*Vitis vinifera* L.) juice. Journal of the Science of Food and Agriculture, 96(2), 664–671. https://doi.org/10.1002/jsfa.7138
- Huang, X., Wu, Y., Zhang, S., Yang, H., Wu, W., Lyu, L., & Li, W. (2022). Variation in Bioactive Compounds and Antioxidant Activity of Rubus Fruits at Different Developmental Stages. *Foods*, 11(8), 1169. https://doi.org/10.3390/foods11081169
- Ivanova, V., Stefova, M., & Chinnici, F. (2010). Determination of the polyphenol contents in Macedonian grapes and wines by standardized spectrophotometric methods. *Journal of the Serbian Chemical Society*, 75(1), 45–59. https://doi.org/10.2298/JSC1001045I
- Jiménez-Monreal, A.M., García-Diz, L., Martínez-Tomé, M., Mariscal, M., & Murcia, M.A. (2009). Influence of Cooking Methods on Antioxidant Activity of Vegetables. *Journal* of Food Science, 74(3), H97–H103. https://doi.org/10.1111/j.1750-3841.2009.01091.x
- Johnson, J., Collins, T., Walsh, K., & Naiker, M. (2020). Solvent extractions and spectrophotometric protocols for measuring the total anthocyanin, phenols and antioxidant content in plums. *Chemical Papers*, 74(12), 4481–4492. https://doi.org/10.1007/s11696-020-01261-8
- Johnson, M. H., & Gonzalez de Mejia, E. (2012). Comparison of Chemical Composition and Antioxidant Capacity of Commercially Available Blueberry and Blackberry Wines in Illinois. *Journal of Food Science*, 77(1), C141–C148. https://doi.org/10.1111/j.1750-3841.2011.02505.x
- Joyanes, M., & Lema, L. (2006). Criteria for Optimizing Food Composition Tables in Relation to Studies of Habitual Food Intakes. *Critical Reviews in Food Science and Nutrition*, 46(4), 329–336. https://doi.org/10.1080/10408390600688156
- Kalt, W., Cassidy, A., Howard, L. R., Krikorian, R., Stull, A. J., Tremblay, F., & Zamora-Ros, R. (2020). Recent Research on the Health Benefits of Blueberries and Their Anthocyanins. *Advances in Nutrition*, 11(2), 224–236. https://doi.org/10.1093/advances/nmz065
- Kassara, S., Norton, E. L., Mierczynska-Vasilev, A., Lavi Sacks, G., & Bindon, K. A. (2022). Quantification of protein by acid hydrolysis reveals higher than expected concentrations in red wines: Implications for wine tannin concentration and colloidal stability. *Food Chemistry*, 385, 132658. https://doi.org/10.1016/j.foodchem.2022.132658
- Lee, J., Dossett, M., & Finn, C. E. (2012). Rubus fruit phenolic research: The good, the bad, and the confusing. *Food Chemistry*, 130(4), 785–796. https://doi.org/10.1016/j.foodchem.2011.08.022
- Leong, S. Y., & Oey, I. (2012). Effects of processing on anthocyanins, carotenoids and vitamin C in summer fruits and vegetables. *Food Chemistry*, 133(4), 1577–1587. https://doi.org/10.1016/j.foodchem.2012.02.052
- Li, J., Shi, C., Shen, D., Han, T., Wu, W., Lyu, L., & Li, W. (2022). Composition and Antioxidant Activity of Anthocyanins and Non-Anthocyanin Flavonoids in Blackberry

from Different Growth Stages. *Foods*, 11(18), 2902. https://doi.org/10.3390/foods11182902

- Li, W.-L., Wu, W.-L., Zhang, C.-H., Lyu, L.-F., Wang, X.-M., & Shu, H.-R. (2012). He status of industry development and scientific research of blackberry (Rubus spp.) in the world, with a prospect in China. *Journal of Plant Resources and Environment*, 21(3), 105–115. https://www.cabdirect.org/cabdirect/abstract/20123334658
- Lillo, A., Carvajal-Caiconte, F., Núñez, D., Balboa, N., & Alvear Zamora, M. (2016). Cuantificación espectrofotométrica de compuestos fenólicos y actividad antioxidante en distintos berries nativos del Cono Sur de América. *Revista de Investigaciones Agropecuarias*, 42(2), 168–174. http://www.scielo.org.ar/scielo.php?script=sci\_abstract&pid=S1669-23142016000200009
- López-Legarda, X., Taramuel-Gallardo, A., Arboleda-Echavarría, C., Segura-Sánchez, F., & Restrepo-Betancur, L.F. (2017). Comparison of methods using sulfuric acid for determination of total sugars. *Revista Cubana De Química*, 29(2), 180–198. http://scielo.sld.cu/pdf/ind/v29n2/ind02217.pdf
- Lykke, A.M., & Padonou, E.A. (2019). Carbohydrates, proteins, fats and other essential components of food from native trees in West Africa. *Heliyon*, 5(5), e01744. https://doi.org/10.1016/j.heliyon.2019.e01744
- Maragò, E., Michelozzi, M., Calamai, L., Camangi, F., & Sebastiani, L. (2016). Antioxidant properties, sensory characteristics and volatile compounds profile of apple juices from ancient Tuscany (Italy) apple varieties. *European Journal of Horticultural Science*, 81(5), 255–263. https://doi.org/10.17660/eJHS.2016/81.5.4
- Martínez-Cruz, N. del S., Arévalo-Niño, K., Verde-Star, M. J., Rivas-Morales, C., Oranday-Cárdenas, A., Núñez-González, Ma. A., & Morales-Rubio, Ma. Eufemia. (2011). Antocianinas y actividad anti radicales libres de *Rubus adenotrichus* Schltdl (zarzamora). *Revista Mexicana de Ciencias Farmacéuticas*, 42(4), 66–71.
- Milošević, T., Milošević, N., Glišić, I., & Mladenović, J. (2012). Fruit quality attributes of blackberry grown under limited environmental conditions. *Plant, Soil and Environment*, 58(7), 322–327. https://doi.org/10.17221/33/2012-PSE
- Minatel, I. O., Borges, C. V., Ferreira, M. I., Gomez, H. A. G., Chen, C.-Y. O., & Lima, G. P.
  P. (2017). Phenolic Compounds: Functional Properties, Impact of Processing and Bioavailability. In M. P.-T. and M. del R. G.-M. Marcos Soto-Hernandez (Ed.), *Phenolic Compounds - Biological Activity* (pp. 1–25). InTech. https://doi.org/10.5772/66368
- Morata, A., López, C., Tesfaye, W., González, C., & Escott, C. (2019). Anthocyanins as Natural Pigments in Beverages. In Alexandru Mihai Grumezescu and Alina Maria Holban (Ed.), *Value-Added Ingredients and Enrichments of Beverages* (Vol. 14, pp. 383–428). Elsevier. https://doi.org/10.1016/B978-0-12-816687-1.00012-6
- Mudnic, I., Modun, D., Rastija, V., Vukovic, J., Brizic, I., Katalinic, V., Kozina, B., Medic-Saric, M., & Boban, M. (2010). Antioxidative and vasodilatory effects of phenolic acids in wine. *Food Chemistry*, 119(3), 1205–1210. https://doi.org/10.1016/j.foodchem.2009.08.038
- Mulero, J., Zafrilla, P., Cayuela, J.M., Martínez-Cachá, A., & Pardo, F. (2011). Antioxidant Activity and Phenolic Compounds in Organic Red Wine Using Different Winemaking Techniques. *Journal of Food Science*, 76(3), C436–C440. https://doi.org/10.1111/j.1750-3841.2011.02104.x
- Murador, D. C., Mercadante, A. Z., & de Rosso, V. V. (2016). Cooking techniques improve the levels of bioactive compounds and antioxidant activity in kale and red cabbage. *Food Chemistry*, 196, 1101–1107. https://doi.org/10.1016/j.foodchem.2015.10.037
- Mustafa, A. M., Angeloni, S., Abouelenein, D., Acquaticci, L., Xiao, J., Sagratini, G., Maggi, F., Vittori, S., & Caprioli, G. (2022). A new HPLC-MS/MS method for the simultaneous determination of 36 polyphenols in blueberry, strawberry and their commercial products and determination of antioxidant activity. *Food Chemistry*, 367, 130743. https://doi.org/10.1016/j.foodchem.2021.130743

- Pantoja-Chamorro, A. L., Hurtado-Benavides, A. M., & Martínez-Correa, H. A. (2017). Evaluación del Rendimiento, Composición y Actividad Antioxidante de Aceite de Semillas de Mora (*Rubus glaucus*) Extraído con CO2 Supercrítico. *Información Tecnológica*, 28(1), 35–46. https://doi.org/10.4067/S0718-07642017000100005
- Parmenter, B. H., Croft, K. D., Hodgson, J. M., Dalgaard, F., Bondonno, C. P., Lewis, J. R., Cassidy, A., Scalbert, A., & Bondonno, N. P. (2020). An overview and update on the epidemiology of flavonoid intake and cardiovascular disease risk. *Food & Function*, 11(8), 6777–6806. https://doi.org/10.1039/D0F001118E
- Pellegrini, N., Miglio, C., Del Rio, D., Salvatore, S., Serafini, M., & Brighenti, F. (2009). Effect of domestic cooking methods on the total antioxidant capacity of vegetables. *International Journal of Food Sciences and Nutrition*, 60(sup2), 12–22. https://doi.org/10.1080/09637480802175212
- Pérez Grana, R. (2013). Exactitud de las tablas de composición de alimentos en la determinación de nutrientes. *Sanidad Militar*, 69(2), 102–111. https://doi.org/10.4321/S1887-85712013000200008
- Robinson, J. A., Bierwirth, J. E., Greenspan, P., & Pegg, R.B. (2020). Blackberry polyphenols: Composition, quantity, and health impacts from in vitro and in vivo studies. *Journal of Food Bioactives*, 9. https://doi.org/10.31665/JFB.2020.9217
- Rodríguez-Pérez, M.A., Hernández Rojas, L. A., Madrigal Ambriz, L.V., García, D.E., Vázquez Galindo, J., & Velasco Villalpando, S. (2010). Evaluación de antocianinas de la zarzamora (Rubus eubatus) en fruta y en conserva. (UANL. Universidad de Guanajuato, Ed.; pp. FH131–FH139). XII Congreso Nacional de Ciencia y Tecnología de Alimentos. https://docplayer.es/18363273-Xii-congreso-nacional-de-ciencia-ytecnologia-de-alimentos-jueves-27-y-viernes-28-de-mayo-de-2010-guanajuatogto.html
- Salvador Badui Dergal. (2006). *Química de los Alimentos*. (Quintanar Duarte Enrique, Ed.; Cuarta edición, Vol. 1). Addison Wesley. https://fcen.uncuyo.edu.ar/upload/librobadui200626571.pdf
- Sánchez Trujillo, A. F. (2013). Fermentación de Autumn Bliss para la elaboración de Vino de Frambuesa. [Tesis Químico en Alimentos, Universidad Autónoma del Estado de México]. http://ri.uaemex.mx/handle/20.500.11799/14125
- Santacruz Cifuentes, L. A. (2011). Análisis químico de antocianinas en frutos silvestres colombianos. [Maestría en Ciencias Química, Universidad Nacional de Colombia]. https://repositorio.unal.edu.co/handle/unal/8678
- Schulz, M., Seraglio, S. K. T., Della Betta, F., Nehring, P., Valese, A. C., Daguer, H., Gonzaga, L. V., Costa, A. C. O., & Fett, R. (2019). Blackberry (*Rubus ulmifolius* Schott): Chemical composition, phenolic compounds and antioxidant capacity in two edible stages. *Food Research International*, 122, 627–634. https://doi.org/10.1016/j.foodres.2019.01.034
- Skrede, G., Wrolstad, R. E., & Durst, R. W. (2000). Changes in Anthocyanins and Polyphenolics During Juice Processing of Highbush Blueberries (*Vaccinium* corymbosum L.). Journal of Food Science, 65(2), 357–364. https://doi.org/10.1111/j.1365-2621.2000.tb16007.x
- Tomas, M., Toydemir, G., Boyacioglu, D., Hall, R., Beekwilder, J., & Capanoglu, E. (2015). The effects of juice processing on black mulberry antioxidants. *Food Chemistry*, 186, 277–284. https://doi.org/10.1016/j.foodchem.2014.11.151
- Tomas, M., Toydemir, G., Boyacioglu, D., Hall, R. D., Beekwilder, J., & Capanoglu, E. (2017). Processing black mulberry into jam: effects on antioxidant potential and *in vitro* bioaccessibility. *Journal of the Science of Food and Agriculture*, 97(10), 3106–3113. https://doi.org/10.1002/jsfa.8152
- Vergara, M. F., Vargas, J., & Acuña, J. F. (2016). Physicochemical characteristics of blackberry (*Rubus glaucus* Benth.) fruits from four production zones of Cundinamarca, Colombia. *Agronomía* Colombiana, 34(3), 336–345. https://doi.org/10.15446/agron.colomb.v34n3.62755

,

Recibido:	Wagner, H., & Bladt, S. (1996). Plant Drug Analysis (Second Edition, Vol. 1). Springer Berlin
30/octubre/2022	Heidelberg. https://doi.org/10.1007/978-3-642-00574-9
	Wu, X., Beecher, G. R., Holden, J. M., Haytowitz, D. B., Gebhardt, S. E., & Prior, R. L. (2006).
	Concentrations of Anthocyanins in Common Foods in the United States and
<b>Aceptado:</b> 1/junio/2023	Estimation of Normal Consumption. Journal of Agricultural and Food Chemistry,
	54(11), 4069–4075. https://doi.org/10.1021/jf0603001
	Yousef, G. G., Brown, A. F., Funakoshi, Y., Mbeunkui, F., Grace, M. H., Ballington, J. R.,
	Loraine, A., & Lila, M. A. (2013). Efficient Quantification of the Health-Relevant
	Anthocyanin and Phenolic Acid Profiles in Commercial Cultivars and Breeding
	Selections of Blueberries (Vaccinium spp.). Journal of Agricultural and Food
	<i>Chemistry</i> , <i>61</i> (20), 4806–4815. https://doi.org/10.1021/jf400823s
	Zafra Rojas, Q. Y. (2019). Valorización de los subproductos del procesamiento de la
	zarzamora (Rubus fruticosus), por su contenido en antioxidantes y fibra dietética.
	[Tesis de Doctorado, Universidad Autónoma del Estado de Hidalgo].
	http://dgsa.uaeh.edu.mx:8080/bibliotecadigital/handle/231104/2675
	Zannou, O., & Koca, I. (2022). Greener extraction of anthocyanins and antioxidant activity
	from blackberry ( <i>Rubus</i> spp) using natural deep eutectic solvents. <i>LWT</i> , 158, 113184.

Trom blackberry (*Rubus* spp) using natural deep eutectic solvents. *LW1*, 158, 113184. https://doi.org/10.1016/j.lwt.2022.113184
Zhishen, J., Mengcheng, T., & Jianming, W. (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chemistry*, 64(4), 555–559. https://doi.org/10.1016/S0308-8146(98)00102-2